

Chronic Kidney Disease, Molecular Insight and Its Herbal Modulation

Afsheen Fatima¹, Asma Imran Ansari³, Farhan Ali², Farzana Mahdi³, Shilpa Sharma⁴, Sher Ali^{5,*}

¹Department of Biotechnology, Era University, Lucknow, UP, India

²Hospitalist, Internal Medicine, Yakima Valley, Memorial Hospital, WA, USA

³Department of Personalized & Molecular Medicine, Era University, Lucknow, UP, India

⁴Department of Paediatrics, All India Institutes of Medical Science, New Delhi, India

⁵Vice Chancellor Office's, Era University, Lucknow, UP, India

Abstract

The human body is equipped with a natural system for internal cleansing and maintenance, with the kidneys serving as the primary organs responsible for filtering waste and maintaining fluid balance. However, modern lifestyle choices including poor diet, excessive intake of harmful beverages, and substance abuse can compromise kidney function. Two major types of kidney diseases are recognized: acute kidney injury (AKI) and chronic kidney disease (CKD). AKI, or acute renal failure, involves a sudden decline in kidney function and is often reversible. In contrast, CKD, also known as chronic renal failure, progresses slowly over at least three months and may lead to irreversible damage if remains untreated. Additional renal conditions include Glomerulonephritis, Polycystic Kidney Disease, Kidney Stones, UTIs, Lupus Nephritis, and nephropathies linked to diabetes and hypertension. While allopathic medications are widely prescribed, their side effects often limit long-term use, especially in elderly or immunocompromised individuals. Herbal formulations are gaining attention as supportive therapies due to their multi-targeted action and lower toxicity. This study considers multiple risk factors diet, lifestyle, alcohol and tobacco use, age, gender, ethnicity, and comorbidities when evaluating herbal interventions. Different kidney disorders may require tailored herbal combinations, warranting systematic research to validate efficacy and safety. Identifying and characterizing bioactive compounds through modern pharmacological techniques can strengthen the clinical credibility of these formulations. This paper aims to present a conceptual framework that bridges traditional herbal medicine with scientific validation, encouraging further exploration of plant-based therapies as complementary, or alternative approaches in kidney disease management.

*CORRESPONDENCE:

Sher Ali,
(profsali55@gmail.com)

KEYWORDS:

Chronic Kidney Disease, Herbal medicine, Renal therapeutics, Plant based formulations, Kidney health management..

HOW TO CITE:

Fatima A, Ansari AI, Ali F, Mahdi F, Sharma S, Ali S. Chronic Kidney Disease, Molecular Insight and Its Herbal Modulation. Journal of Academy of Biomedical Sciences. 2025;2(1):41-55

doi: 10.61081/jabs/2v1i108

INTRODUCTION

Chronic kidney disease (CKD) is no longer an uncommon disorder; it is a looming global health challenge. It already ranks among the top ten causes of death and is projected to reach fifth place by 2040, creating major clinical and economic.^{1,2} The pathophysiology of CKD is complex and multifactorial, common features include scarring of the glomeruli (glomerulosclerosis), shrinking of kidney tubules (tubular atrophy), tissue fibrosis and chronic inflammation.³ At the molecular level, several signalling pathways such as NF- κ B, TGF- β /Smad, MAPK, PI3K/Akt, and Nrf2 drive oxidative stress, inflammation, cell death, and fibrosis. Other contributing factors include overactivation of the renin-

angiotensin-aldosterone system (RAAS), mitochondrial damage, and metabolic imbalances.⁴ Recent research points to the gut-kidney axis as a key player in systemic inflammation and toxin clearance.⁵ Beyond environmental and metabolic factors, genetics plays a crucial role in CKD susceptibility and progression. Genome-wide association studies (GWAS), and candidate gene analyses have revealed multiple genetic loci associated with renal function and eGFR,⁶ with more than 600 genes implicated in kidney physiology and pathology.⁷ Some forms of CKD are caused by single-gene (monogenic) defects, such as Alport syndrome (mutations in *COL4A3*, *COL4A4*, *COL4A5*), autosomal dominant polycystic kidney disease (*PKD1*, *PKD2*), and autosomal

recessive polycystic kidney disease (*PKHD1*), which affect structural proteins or ciliary signalling. More commonly, CKD is influenced by multiple genes (polygenic). Variants in genes like *UMOD*, *APOL1*, *SHROOM3*, and *SLC7A9* increase susceptibility.⁸ Notably, *APOL1* alleles (G1, G2) confer markedly higher risk for focal segmental glomerulosclerosis (FSGS) and hypertensive nephropathy in individuals of African ancestry. Likewise, variations in RAAS genes (*ACE*, *AGT*, *AGTR1*) and oxidative stress regulators can worsen CKD in individuals with diabetes or hypertension.⁹ Epigenetic mechanisms such as DNA methylation, histone modifications, and noncoding RNAs further modulate disease pathways, adding another regulatory layer.¹⁰ Current therapeutic strategies focus on slowing disease progression and managing complications through agents like ACE inhibitors, angiotensin receptor blockers (ARBs), and SGLT2 inhibitors.¹¹ While, these drugs reduce proteinuria and delay progression, they cannot reverse structural damage and often cause adverse effects such as hyperkalaemia and hypotension. In advanced stages, dialysis or kidney transplantation remains the only option, both associated with high costs, and significant psychosocial burdens.¹² This underscores the urgent need for safe, cost-effective, multi-targeted therapeutic alternatives. Herbal medicine, rooted in traditional systems such as Ayurveda and Traditional Chinese Medicine (TCM), offers potential solutions. Many plants traditionally used for kidney disorders are now validated by modern research, showing antioxidant, anti-inflammatory, antifibrotic, and immunomodulatory activities.¹³ Molecular studies reveal that several botanicals can modulate signalling pathways implicated in CKD. For example, curcumin from *Curcuma longa* activates Nrf2, enhancing antioxidant gene expression (*HO-1*, *GCLC*, *GCLM*, *NQO1*, *GST*) and reducing oxidative stress, inflammation, and fibrosis, in experimental models.¹⁴ *Astragalus membranaceus*, and its active compound astragaloside IV inhibit TGF- β /Smad signalling, downregulating fibrosis-related genes (*COL1A1*, *FN1*, α -*SMA*) and restoring *CDH1*, thereby preventing epithelial-to-mesenchymal transition (EMT) and renal scarring.¹⁵ Other botanicals, including *Salvia miltiorrhiza* and berberine, though less studied at the gene-specific level, exhibit significant nephroprotective effects. They suppress TGF- β /Smad and NF- κ B signalling while activating AMPK, thereby reducing inflammation and fibrosis.¹⁶ These multi-targeted actions position herbal medicines as promising adjunct therapies. However, challenges remain, including variability in phytochemical content, lack of standardization, herb-drug interactions, and scarcity of large-scale randomized clinical trials. Emerging omics technologies genomics, transcriptomics, proteomics, and metabolomics combined with network pharmacology and bioinformatics, offer new opportunities to map herb-gene interactions and optimize therapeutic strategies.¹⁷ Identifying key molecular targets, pathways, and gene expression profiles modulated by phytochemicals will not only validate traditional practices but also accelerate discovery of novel nephroprotective

agents. In conclusion, herbal therapeutics hold significant potential in CKD management by attenuating oxidative stress, inflammation, and fibrosis through modulation of critical signalling networks. Bridging the gap between traditional practices and molecular evidence through robust clinical trials and advanced mechanistic research is critical for integrating herbal medicine into evidence-based nephrology.

MONOGENIC KIDNEY DISEASES

Cystic Kidney Diseases

Autosomal Dominant Polycystic Kidney Disease (ADPKD)

Autosomal Dominant Polycystic Kidney Disease (ADPKD) is one of the most common inherited kidney disorders, characterized by the progressive development of renal cysts (Figure 1), leading to enlarged kidneys and eventual renal failure.¹⁸ The molecular foundation of ADPKD lies in mutations affecting two critical genes: *PKD1* (85–90% of cases) and *PKD2* (10–15% of cases), which encode polycystin-1 (*PC1*) and polycystin-2 (*PC2*) proteins, respectively.¹⁸ These polycystins form a functional complex primarily located in the primary cilia of renal epithelial cells, where they serve as mechanosensors responding to fluid flow and mediate crucial calcium signalling pathways.¹⁹ Loss-of-function mutations in *PKD1* or *PKD2* impair calcium signalling, leading to altered expression of calcium-regulated genes, including those controlling proliferation (*CCND1*, *cyclin D1*), apoptosis (*BCL2*, *CASP3*), and fluid transport (*CFTR*).²⁰ This genetic disruption creates a cascade of transcriptional changes that promote cyst initiation and expansion. Hyperactivation of cAMP-dependent pathways in ADPKD results from increased expression of genes encoding adenylyl cyclase isoforms (*ADCY6*) and reduced expression of phosphodiesterase genes (*PDE1*), elevating intracellular cAMP levels.²¹ This upregulates PKA-regulated transcription factors such as CREB, which enhance expression of proliferative genes (*MYC*, *FOS*, *EGR1*), chloride transporters (*CFTR*), and secretory machinery genes, accelerating cyst growth. Similarly, aberrant activation of the mTOR pathway reflects dysregulation of *TSC2* (tuberin), normally stabilized by *PC1*. Loss of *PC1* function decreases *TSC2* activity, increasing expression of genes involved in protein synthesis (*RPS6KB1*, *S6 kinase*) and cell growth regulators (*EIF4EBP1*), further driving cystic expansion.²²

Autosomal Recessive Polycystic Kidney Disease (ARPKD)

Autosomal recessive polycystic kidney disease (ARPKD) is a severe hereditary disorder typically presenting in infancy or childhood characterized by cystic dilation of the collecting ducts and congenital hepatic fibrosis.²³ The molecular

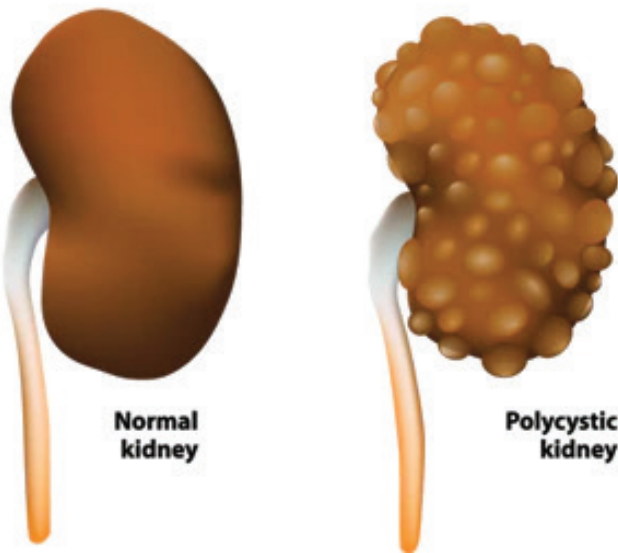


Figure 1: Morphologies of normal and polycystic kidney.

foundations and cellular mechanisms of ARPKD centre on defects in primary cilia and associated signalling pathways in renal tubular epithelial cells. Most ARPKD cases result from mutations in the *PKHD1* gene, which encodes fibrocystin/polyductin (*FPC*), a large transmembrane protein.²⁴ *FPC* is predominantly located in the primary apical cilia and basal bodies of kidney epithelial cells, as well as cholangiocytes in the liver. Primary cilia are non-motile, microtubule-based organelles that sense mechanical and chemical stimuli in the renal tubule lumen and regulate crucial signalling pathways involved in cell proliferation and differentiation.²⁵ Loss of function in *FPC* disrupts the structural integrity and signalling functions of the primary cilium. This leads to impaired regulation of cell growth, differentiation, and tubulogenesis, resulting in abnormal and uncontrolled dilatation of the collecting ducts. *FPC* interacts indirectly with other PKD proteins (*PC1* and *PC2*, involved in ADPKD), and there is evidence that PKD proteins form a functional ciliary complex, sharing downstream effectors such as the mTOR, MAPK/ERK, cAMP/PKA, and calcium signalling pathways.²⁶ Ciliary dysfunction alters calcium influx, disrupting calcium-dependent signalling and affecting pathways governing cell proliferation and fluid secretion. Abnormal regulation of adenylate cyclase and phosphodiesterase lead to elevated cAMP levels, which, via PKA activation, promotes cystic cell proliferation and fluid secretion into cysts. Defects in sodium and chloride channels (*ENaC*, *CFTR*) and aquaporins (*AQP2*) contribute to disturbed fluid movement and cyst expansion.²⁷ TGF- β , EGFR, and VEGF signalling are implicated in fibrosis and abnormal tissue transformation in ARPKD.²⁸

Nephronophthisis

Nephronophthisis (NPHP) is an autosomal recessive ciliopathy

and the leading genetic cause of end-stage renal disease in children and young adults. It is characterized by chronic tubulointerstitial fibrosis, and progressive cyst formation that culminates in kidney failure within the first three decades of life. The disease arises from mutations in more than 26 genes (*NPHP1–26*), most of which encode proteins critical for primary cilium function, with *NPHP1* mutations accounting for 21–25% of cases, while other individual genes contribute less than 3% each. Approximately, 70% of cases remain genetically unresolved, highlighting the likelihood of additional unidentified genes.²⁹ The protein products of *NPHP* genes, termed nephrocystins, are primarily localized to the transition zone or inversion compartment of primary cilia, centrosomes, or components of intraflagellar transport complexes. They operate in modules such as *NPHP14–8*, *NPHP2–3–9–16*, *NPHP5–6*, and the MKS module. Disruption of these ciliary structures leads to multiple signalling defects, including abnormal Hedgehog (Hh) signalling marked by increased Gli3 repressor protein levels, which can be partially rescued by the Hh agonist purmorphamine. Additionally, defective inversin skews the Wnt pathway toward canonical signalling at the expense of non-canonical planar cell polarity (PCP) signalling, thereby impairing epithelial polarity. *NPHP* pathogenesis involves dysregulated cAMP/PKA signalling, where impaired ciliary control reduces basal cAMP levels essential for Gli regulation and cellular homeostasis. Emerging evidence also implicates defective DNA damage response mechanisms, leading to genomic instability and cell cycle abnormalities, suggesting that *NPHP* is not a solely ciliary disease but also involves broader cellular pathway.^{30,31}

Medullary Cystic Kidney Disease (MCKD)

Medullary Cystic Kidney Disease (MCKD) is an autosomal dominant inherited disorder. It is characterized by progressive tubulointerstitial fibrosis and the formation of cysts in the renal medulla. This leads to adult-onset chronic kidney disease and eventually progresses to end-stage renal failure. MCKD is closely related to nephronophthisis (NPH) and together, they are sometimes referred to as the nephronophthisis–medullary cystic kidney disease complex due to overlapping clinical and genetic features.³² MCKD is primarily caused by mutations in two genes: *MUC1* (*MCKD1*, chromosome1), and *UMOD* (*MCKD2*, chromosome 16p12). The *UMOD* gene encodes uromodulin (Tamm-Horsfall protein), which is the most abundant protein in normal urine and is expressed by the thick ascending limb cells of the loop of Henle in the kidney. Mutations in *UMOD* lead to the accumulation of misfolded uromodulin in the endoplasmic reticulum (ER) of renal tubular epithelial cells, triggering ER stress, apoptosis, and progressive tubular damage and fibrosis. This process also results in decreased urinary excretion of uromodulin, consistent with MCKD's clinical findings. Specific exons of *UMOD* (notably exons 4 and 5) are important for its calcium-binding and folding properties. Mutations disrupt protein structure and trafficking, exacerbating ER stress

and damage. Chronic accumulation of mutant uromodulin in the ER induces apoptosis and a scarring process, leading to tubulointerstitial injury and renal fibrosis. The disease process is not linked to ciliary dysfunction, unlike many other cystic kidney diseases, since *UMOD* is not expressed in the cilia of tubular epithelial cells. Inflammation and immune mechanisms may also be implicated, as protein aggregates can stimulate antibody formation and cellular infiltration, enhancing the risk of progressive renal insufficiency and scarring. Unlike polycystic kidney diseases (PKD), MCKD cysts are usually fewer, smaller, limited to the corticomedullary junction, and are secondary to underlying tubular atrophy and interstitial fibrosis rather than primary cyst formation.³³

GLOMERULAR DISEASES

1. Alport Syndrome

Alport syndrome is a hereditary kidney disorder caused by mutations in *COL4A3*, *COL4A4*, and *COL4A5*, which encode the $\alpha3$, $\alpha4$, and $\alpha5$ chains of type IV collagen, essential for the structural integrity of the glomerular basement membrane (GBM).³⁴ These mutations disrupt the assembly of the $\alpha3\alpha4\alpha5$ (IV) collagen heterotrimer. This leads to thinning of the glomerular basement membrane (GBM), splitting of the lamina densa and a characteristic basket-weave appearance. As a result, affected individuals develop progressive glomerulosclerosis, kidney failure, sensorineural hearing loss, and ocular abnormalities.³⁵ Abnormal collagen assembly also alters extracellular matrix remodelling, causing aberrant laminin deposition and persistence of immature isoforms, which correlate with podocyte injury. Furthermore, changes in integrin signalling, particularly overexpression of $\alpha\beta6$, activate profibrotic pathways such as TGF- β and increase matrix metalloproteinase activity (*MMP2*, *MMP9*, *MMP14*), promoting mesangial migration and fibrosis. Proteomic studies also reveal mitochondrial dysfunction and impaired energy metabolism through *PPAR α* pathway downregulation, indicating metabolic dysregulation as a potential therapeutic target.³⁶

2. Fabry Disease

Fabry disease is a rare X-linked lysosomal storage disorder caused by mutations in the *GLA* gene, which encodes the enzyme α -galactosidase A (*α -GalA*). Enzyme deficiency leads to progressive accumulation of glycosphingolipids, primarily globotriaosylceramide (Gb3) and its derivative lyso-Gb3, within lysosomes. This buildup occurs across multiple tissues, disrupting normal cellular function and initiating complex pathogenic cascades. Gb3 accumulation results in lysosomal dysfunction and impaired autophagy, contributing to defective organelle turnover and mitochondrial injury.³⁷ These changes exacerbate oxidative stress and mitochondrial dysfunction, promoting reactive oxygen species (ROS) production and oxidative damage, partly through RIPK3-dependent mechanisms. Furthermore, inflammatory

pathways are activated, with macrophage polarization, elevated cytokine expression, and activation of the NF- κ B pathway, leading to chronic tissue injury and fibrosis.³⁸ Additionally, Gb3-driven energy depletion alters metabolic signalling, activating AMP-activated protein kinase (AMPK) and inhibiting mTORC1, impairing autophagy and shifting cellular metabolism from fatty acid oxidation to glycolysis.³⁹ Collectively, these molecular events lipid storage, oxidative stress, immune activation, and metabolic dysregulation drive the multisystemic manifestations of Fabry disease.

3. Pierson Syndrome

Pierson syndrome is a rare autosomal recessive disorder caused primarily by mutations in the *LAMB2* gene, which encodes the laminin $\beta2$ subunit, an essential component of basement membranes, particularly the glomerular basement membrane (GBM) in the kidney and ocular structures.⁴⁰ Laminin $\beta2$ normally forms part of the $\alpha5\beta2\gamma1$ heterotrimer, crucial for GBM stability and podocyte adhesion. Loss or dysfunction of laminin $\beta2$ disrupts extracellular matrix organization, integrin signalling, and slit diaphragm integrity, resulting in congenital nephrotic syndrome, ocular abnormalities, and, in some cases, neuromuscular defects. Functionally connected genes, including *NPHS1*, *NPHS2*, *COL4A3*, *COL4A5*, *PLCE1*, and *AGRN*, contribute to glomerular development and podocyte function, explaining overlapping clinical features with other congenital nephrotic syndromes.⁴⁰ The severity of Pierson syndrome correlates with *LAMB2* mutation type: truncating variants typically cause complete loss of protein, whereas missense or compound heterozygous mutations lead to partial function and variable phenotypes. Overall, the disorder reflects a fundamental defect in basement membrane assembly, affecting GBM integrity, neuromuscular junctions, and ocular structure maintenance.⁴¹

4. Congenital Nephrotic Syndrome (Finnish Type) (CNF)

It is a severe autosomal recessive kidney disorder characterized by massive proteinuria and edema, typically manifesting in utero or shortly after birth. The condition results primarily from mutations in the *NPHS1* gene, which encodes nephrin, a transmembrane protein crucial for the structure and function of the glomerular slit diaphragm (SD), the main size-selective filtration barrier in the kidney. Nephrin belongs to the immunoglobulin superfamily and forms a zipper-like structure between podocyte foot processes. Its absence or dysfunction disrupts SD integrity, causing increased glomerular permeability and massive proteinuria. Ultrastructural changes include podocyte foot process effacement and disappearance of slit diaphragms. In Finland, CNF is associated with two founder *NPHS1* mutations Fin-major and Fin-minor which account for most cases. While *NPHS1* mutations define CNF, other podocyte-related genes (e.g., *NPHS2*, *ACTN4*) can cause similar but distinct congenital

nephrotic syndromes. Overall, *NPHS1* mutations and nephrin deficiency are the molecular hallmark of CNF, leading to disruption of the glomerular filtration barrier and severe proteinuric nephropathy.⁴²

5. Denys-Drash syndrome (DDS) and Frasier syndrome (FS)

These are rare disorders caused by mutations in the *WT1* gene on chromosome 11p13, which encodes a zinc-finger transcription factor essential for kidney and gonadal development. *WT1* regulates genes critical for nephron differentiation, podocyte function, and gonadal morphogenesis. DDS is most commonly associated with missense mutations in exons 8 and 9 within the DNA-binding zinc finger domains, particularly the p.Arg394Trp variant, leading to dominant-negative disruption of *WT1*'s transcriptional regulation and resulting in diffuse mesangial sclerosis, gonadal dysgenesis, and a high risk of Wilms tumour. Conversely, FS typically arises from splice site mutations in exon 9 that alter the ratio of *WT1* isoforms containing or lacking the KTS sequence, shifting the normal 2:1 (+KTS/–KTS) balance to approximately 1:2.⁴³ This isoform imbalance affects podocyte gene expression and gonadal development, causing progressive nephropathy (commonly focal segmental glomerulosclerosis), gonadal dysgenesis, and an increased risk of gonadoblastoma but rarely Wilms tumour. Both syndromes disrupt pathways involved in slit diaphragm integrity, mesenchymal-epithelial transition, and glomerular development, intersecting with genes such as *NPHS1*, *NPHS2*, *PAX2*, and *CD2AP*.⁴⁴

6. Thin Basement Membrane Nephropathy (TBMN)

TBMN is primarily associated with heterozygous mutations in *COL4A3* and *COL4A4*, which encode the $\alpha 3$ and $\alpha 4$ chains of type IV collagen essential components of the mature glomerular basement membrane (GBM).⁴⁵ Normally, the GBM undergoes a developmental transition from an $\alpha 1$: $\alpha 2$ collagen IV network to an $\alpha 3$: $\alpha 4$: $\alpha 5$ network, providing structural integrity and selective permeability. In TBMN, pathogenic variants most often single-nucleotide substitutions unique to families disrupt assembly of the $\alpha 3\alpha 4\alpha 5$ collagen IV network, causing diffuse GBM thinning and persistent microscopic hematuria.⁴⁶ Beyond the core *COL4A3*/*COL4A4* mutations, modifying variants in genes regulating podocyte cytoskeleton and basement membrane interactions (e.g., *MYO1E*, *LAMA5*, *NPHS2*, *ITGB4*) have been reported to influence disease severity and progression to chronic kidney disease.⁴⁷ Disruption of collagen IV architecture also alters GBM interactions with laminins and nidogens, impacting extracellular matrix signalling and podocyte adhesion.⁴⁵ Overall, TBMN reflects a primary defect in collagen IV network assembly, with secondary involvement of podocyte and extracellular matrix pathways modulating phenotypic variability.

TUBULAR AND ELECTROLYTE DISORDERS

1. Bartter Syndrome

Bartter syndrome comprises a group of rare inherited renal tubulopathies characterized by defective salt reabsorption in the thick ascending limb (TAL) of the loop of Henle, leading to hypokalaemia, metabolic alkalosis, and secondary hyperreninemia.⁴⁸ The disorder is genetically heterogeneous, involving mutations in genes encoding key transporters and channels in the TAL. Type 1 results from mutations in *SLC12A1*, encoding the Na-K-2Cl cotransporter (*NKCC2*), essential for sodium chloride reabsorption. Type 2 involves *KCNJ1* mutations affecting potassium channels, disrupting potassium recycling required for *NKCC2* function. Type 3 is caused by *CLCNKB* mutations impairing basolateral chloride exit, while Type 4a results from *BSND* mutations encoding barttin, an accessory subunit for ClC-Ka/Kb channels, and is associated with sensorineural deafness. Combined *CLCNKA* and *CLCNKB* mutations produce a similar phenotype (Type 4b). Type 5 involves *MAGED2* mutations (X-linked), affecting *NKCC2* trafficking, causing transient antenatal Bartter syndrome.⁴⁹ A CaSR gain-of-function variant mimicking Bartter syndrome further illustrates the regulatory role of calcium signalling in TAL function. Disruption of these pathways causes salt wasting, chronic prostaglandin E2 activation, and polyuria, highlighting how diverse genetic defects converge on impaired TAL salt handling.

2. Gitelman Syndrome (GS)

Gitelman syndrome is an autosomal recessive salt-wasting tubulopathy caused by impaired sodium-chloride reabsorption in the distal convoluted tubule (DCT), clinically characterized by hypokalaemia, hypomagnesemia, metabolic alkalosis, and hypocalciuria.⁵⁰ The primary genetic defect occurs in *SLC12A3*, which encodes the thiazide-sensitive Na-Cl co-transporter (NCC), essential for sodium and chloride reabsorption; its dysfunction leads to salt wasting and the hallmark electrolyte abnormalities.⁵¹ Mutations in *CLCNKB*, encoding the ClC-Kb chloride channel, or in genes indirectly modulating NCC function such as *KCNJ10*, *KCNJ16*, *FXYP2*, *ATP1A1*, and *HNF1B* can produce Gitelman-like phenotypes. Additionally, maternally inherited variants in mitochondrial tRNA genes (*MT-TI*, *MT-TF*) have been implicated in rare Gitelman-like syndromes, likely due to impaired ATP generation required for NCC phosphorylation and transport. These findings underscore that while NCC dysfunction remains the central defect, GS reflects a broader disruption of pathways governing sodium, chloride, potassium, and magnesium handling in the DCT, often involving complex regulatory and metabolic networks.⁵²

3. Cystinuria

Cystinuria is an inherited renal tubular disorder characterized by defective reabsorption of cystine and dibasic amino acids (ornithine, lysine, arginine) in the proximal tubule and small

intestine, leading to excessive urinary cystine excretion and recurrent nephrolithiasis.⁵³ The condition primarily results from mutations in *SLC3A1* and *SLC7A9*, which encode the heavy and light subunits of the heterodimeric amino acid transporter localized on the apical membrane of proximal tubular epithelial cells.⁵⁴ These subunits are essential for renal amino acid reabsorption; pathogenic variants disrupt transporter assembly or function, resulting in cystinuria types A, B, or AB depending on whether *SLC3A1*, *SLC7A9*, or both are mutated.⁵⁵ Since cystine is poorly soluble in urine, elevated levels promote crystal precipitation and stone formation. In rare cases, larger chromosomal deletions involving *SLC3A1* and neighbouring genes such as *PREPL* can lead to syndromic forms with neuromuscular symptoms. Overall, cystinuria exemplifies a disorder of amino acid transport with high clinical relevance for stone disease and potential genetic counselling implications.⁵⁵

POLYGENIC KIDNEY DISEASES

1. Chronic Kidney Disease (CKD)

CKD involves a complex interplay of inflammatory, metabolic, and fibrotic pathways, influenced by many genetic factors. Inflammation is a key driver, with genes such as *ICAM1*, *CXCL8*, and *CCL2* mediating leukocyte infiltration and cytokine-driven injury. Oxidative stress further exacerbates kidney damage, where *NRF2* functions as a central regulator of antioxidant responses and metabolic homeostasis.⁵⁶ Additionally, activation of signalling pathways such as PI3K-Akt and dysregulation of cell cycle and extracellular matrix genes (*CDKN1C*, *COL4A2*, *SERPINE2*) contribute to fibrosis and progressive nephron loss, while downregulation of podocyte-specific genes like *NPHS1* compromises the filtration barrier.⁵⁷ These findings underscore the polygenic nature of CKD, with over 600 implicated genes converging on common molecular networks that drive chronic inflammation, oxidative injury, and fibrogenesis.⁵⁸

2. Diabetic Nephropathy (DN)

Diabetic nephropathy is a progressive microvascular complication of diabetes driven by interlinked pathways involving inflammation, oxidative stress, and fibrosis. Key molecular mechanisms include immune and inflammatory pathways such as complement activation, chemokine signalling, and antigen-antibody interactions, which underline the strong immune-mediated nature of DN.⁵⁹ Hyperglycaemia induces activation of PI3K-Akt and Rap1 signalling pathways, contributing to altered cell survival, proliferation, and metabolism. NF κ B and MAPK signalling cascades, activated by oxidative stress and advanced glycation end-products (AGEs), further amplify inflammatory and apoptotic responses in podocytes and tubular epithelial cells. Among profibrotic mechanisms, the TGF- β 1 pathway plays a central role in driving extracellular matrix (ECM) accumulation and fibrosis, often enhanced by angiotensin-II

and protein kinase C (PKC) signalling. Additionally, epigenetic modifications such as DNA methylation at the *TXNIP* locus mediate metabolic memory; sustaining disease progression even after glycaemic control improves. JAK/STAT signalling, particularly through *STAT3*, also regulates inflammatory cytokines and Notch1-dependent profibrotic gene expression.⁶⁰ Several key genes have been identified as central regulators of DN pathology, including *TYROBP*, *ITGB2*, *CD53*, *IL10RA*, *LAPTM5*, *CD48*, *C1QA*, and *IRF8*. Additional transcription factors like *SPI1*, *HIF1A*, *STAT1*, *KLF5*, *RUNX1*, and *WT1* contribute to podocyte injury, endothelial dysfunction, and progression of renal fibrosis.⁶¹

3. Hypertensive Nephropathy (HN)

It is a progressive kidney disorder driven by sustained high blood pressure, leading to structural and functional damage through a complex network of molecular pathways and genetic regulators. Key mechanisms include proteasome-mediated protein degradation, which alters turnover of regulatory proteins, and dysregulation of Rho GTPase signalling and cytoskeletal organization, impairing podocyte and tubular integrity. Inflammatory signalling pathways such as NF- κ B, IFN- γ , IL-12, and Wnt are upregulated, promoting immune cell infiltration and chronic inflammation, while metabolic derangements involving the tricarboxylic acid (TCA) cycle and amino acid/purine metabolism highlight the role of HN as a metabolic disorder.⁶² Additionally, renal tissue hypoxia and oxidative stress exacerbate injury, triggering redox imbalance and reactive oxygen species accumulation. Principal genes implicated in these pathways include *SORD*, *CUBN*, and albumin, associated with energy metabolism, and *POLR2I*, *POLR2L*, and *POLR2G*, subunits of RNA polymerase II involved in transcriptional regulation and oxidative stress responses. *TRIM31*, an E3 ubiquitin ligase, promotes proteasomal degradation of *MAP3K7*, thereby modulating inflammation. Other significant genes include *NR4A1*, *TNFSF10*, *CX3CR1*, *CXCR4*, *CCL5*, and *ATF3*, which regulate cytokine signalling and immune activation. Epigenetic and non-coding RNA regulation also play critical roles, with microRNAs such as hsa-miR-1248, hsa-miR-200b-5p, and hsa-miR-23b-5p fine-tuning gene expression.⁶³ Functionally, these interactions affect pathways such as proteasome-mediated degradation (*TRIM31*, *MAP3K7*), TCA cycle and metabolic processes, transcriptional regulation and oxidative stress (*POLR2I*, *POLR2L*, *POLR2G*), inflammatory signalling (*NR4A1*, *TNFSF10*, *NF- κ B*, *CCL5*), and cytoskeletal integrity (actin-associated proteins). Collectively, these molecular alterations converge to cause progressive renal fibrosis, glomerular sclerosis, and loss of kidney function in HN.⁶³

4. Lupus Nephritis

Lupus nephritis (LN) is driven by a complex interplay of genetic and molecular factors that promote immune complex deposition, inflammation, and kidney tissue damage. Recent studies have identified key genes and pathways central

to LN pathogenesis. Driver genes such as *CD53*, *TGFBI*, *MS4A6A*, and *HERC6* are implicated in immune activation, macrophage activity, and inflammation, correlating with disease severity and proteinuria. Other upregulated genes, including *C1QA*, *C1QB*, *IFI44L*, *TYROBP*, and *MS4A4A*, participate in complement activation, interferon signalling, and macrophage differentiation, underscoring their role in immune cell infiltration. Polymorphisms in *IRF5*, *IRF7*, *TNFAIP3*, *TNIP1*, and *UBE2L3* regulate type I interferon, NF- κ B, and MAPK pathways, enhancing autoantibody production and inflammatory signaling.⁶⁴ Additionally, epigenetic changes, including DNA methylation affecting genes such as *ITGAL*, *TNFSF7*, and *CD40LG*, further contribute to immune dysregulation. Critical pathways such as complement activation, toll-like receptor (TLR)/MyD88 signalling, and type I interferon responses play dominant roles, while HLA-DRB1 strongly associates with LN susceptibility⁶⁵.

5. IgA Nephropathy (Berger's Disease)

IgA nephropathy (IgAN), or Berger's disease, is characterized by the deposition of IgA antibodies in the glomeruli, leading to inflammation and progressive kidney damage. The disease arises from a complex interplay of immune dysregulation, genetic susceptibility, and abnormal IgA1 glycosylation. Aberrantly glycosylated IgA1 (galactose-deficient IgA1, Gd-IgA1) forms immune complexes with glycan-specific autoantibodies, triggering mesangial deposition and renal injury. Several signalling pathways, including NF- κ B, JAK-STAT, TGF- β , MAPK, and PI3K-Akt, regulate inflammation, immune responses, fibrosis, and cell survival in IgAN. Long noncoding RNAs (lncRNAs) modulate these processes via PI3K/AKT/mTOR, PTEN, Notch, and JNK pathways, influencing immune activation and kidney damage. lncRNAs such as NQO1-DT, RP5-1057120.6, and G21551 have emerged as potential biomarkers and therapeutic targets. Genetically, GWAS studies have revealed multiple susceptibility loci associated with immune regulation (e.g., *TNFSF4*, *TNFSF18*, *REL*, *CD28*, *LY86*, *LYN*, *ETS1*, *IRF8*, *TNFRSF13B*, *FCAR*) and IgA production or clearance (*IGH*, *FCGR3B*). Genes controlling glycosyltransferases are particularly important in mediating the IgA1 glycosylation defect, which has an estimated heritability of 50–70%. Hub genes such as *FOS*, *JUN*, *EGFR*, *SIRT1*, *IGF1*, *HIF1A*, *SOCS3*, *ACTR2*, and *CTNBN1* are implicated in inflammatory responses, cell proliferation, and apoptosis, highlighting key molecular drivers of disease progression.⁶⁶

6. Kidney Stone Disease (Nephrolithiasis)

A multifactorial condition influenced by genetic, metabolic, and environmental factors, with recent studies emphasizing the involvement of specific molecular pathways and genes. Among key genes, *KLK1* (Kallikrein 1) and *MMP10* (Matrix Metalloproteinase-10) have emerged as significant diagnostic markers, showing upregulation in stone-bearing kidneys where they contribute to metabolic regulation, inflammation, immune responses, and extracellular matrix remodelling,

thereby promoting stone formation.⁶⁷ Other genes, including *LCN2*, *IL11*, *PTGS1*, *GPX3*, and *MMD*, are upregulated in Randall's plaque, the nidus for stone anchoring, while genes such as *SLC12A1* and *NALCN* are downregulated, implicating changes in ion transport and tubular function. Variations in *ATP1A1*, which encodes the Na⁺/K⁺-ATPase α 1-subunit, reduce its expression and may promote renal crystal deposition by altering sodium and calcium homeostasis. Broader genetic studies also highlight both monogenic forms, such as mutations causing familial hypomagnesemia with hypercalciuria and nephrocalcinosis, and polygenic risk, suggesting that multiple low-effect alleles modulate susceptibility.⁶⁸ At the molecular level, pathways regulating ion transport, metabolic homeostasis, and oxalate/calcium balance are central to stone pathogenesis. Inflammatory and immune responses, mediated by cytokines and immune cell recruitment, contribute to renal tissue injury and crystal retention, while oxidative stress from reactive oxygen species promotes tubular damage and alters gene expression linked to stone growth. Moreover, signalling cascades such as MAPK, PI3K/Akt, and those associated with type I diabetes are enriched in stone-forming tissues, reflecting their role in stress response, apoptosis, and metabolic dysregulation. Dysregulation of sodium and calcium reabsorption through channels and pumps, including Na⁺/K⁺-ATPase and related transporters, further increases urinary supersaturation, reinforcing the risk of stone formation.⁶⁹

KIDNEY CANCER

Kidney Cancer, also known as renal cancer, is a malignancy that originates in the kidney tissue, with renal cell carcinoma (RCC) being the most common subtype, accounting for nearly 90% of cases. This cancer typically develops in the renal tubules and is often detected incidentally through imaging studies, although advanced stages present with symptoms such as haematuria, flank pain, and unexplained weight loss. RCC pathogenesis involves complex molecular and genetic alterations that promote uncontrolled cell growth, angiogenesis, and immune evasion. A hallmark of RCC is inactivation of the *VHL* (Von Hippel-Lindau) tumour suppressor gene, leading to stabilization of hypoxia-inducible factors (*HIF-1 α* and *HIF-2 α*) under normoxic conditions. This upregulates target genes such as *VEGF* and *PDGF*, driving angiogenesis and tumour growth.⁷⁰ Another critical signalling network in RCC is the PI3K/AKT/mTOR pathway, frequently activated through mutations in mTOR, TSC1, and TSC2, promoting uncontrolled cell proliferation and inhibiting autophagy.⁷¹ Dysregulated angiogenic signalling through VEGF is a major feature of RCC and forms the therapeutic basis for VEGF-targeted tyrosine kinase inhibitors. In addition, *MET* proto-oncogene alterations, often seen in papillary RCC, activate downstream RAS/RAF/MAPK signalling, enhancing tumour progression.⁷⁰ Epigenetic dysregulation is also common, with frequent mutations in chromatin remodelling genes such as *PBRM1*, *BAP1*, and *SETD2*, which affect transcription, DNA repair, and

immune microenvironment; notably, BAP1 loss is associated with aggressive disease, while *PBRM1* mutations influence immunotherapy response.⁷² Furthermore, immune evasion mechanisms, particularly overexpression of *PD-1/PD-L1*, contribute to RCC progression and justify the use of immune checkpoint inhibitors in treatment.⁷³ Collectively, these molecular events illustrate that RCC is driven by an interplay of pathways involving hypoxia signalling, growth factor receptor activation, chromatin remodelling, and immune modulation, which has paved the way for targeted therapies and immunotherapies that have significantly improved patient outcomes.

COMPLEX GENE INTERACTIONS

A comprehensive search in the Online Mendelian Inheritance in Man (OMIM), revealed 18,683 gene entries mapped across the human genome, distributed across all autosomes and sex chromosomes, including 56 genes on the Y chromosome.⁷⁴ Notably, several transcripts such as members of the TTTY family were present in this dataset. Chromosome-specific evaluation indicated that nearly every chromosome contains genes potentially associated with CKD (Table.1.). While these findings do not imply that all 18,000+ genes directly contribute to CKD, they highlight the extensive genetic complexity of the disease. This broad association likely reflects the involvement of numerous biological pathways that regulate renal development, glomerular filtration, tubular transport, immune modulation, and fibrosis. Many of these pathways operate in a stage-specific and context-dependent manner to maintain kidney function. The large gene count may also arise from pleiotropic effects, wherein variants in genes implicated in metabolic, inflammatory, or vascular processes exert indirect effects on CKD susceptibility. Although the exact number of genes actively driving CKD remains uncertain, current genomic and transcriptomic studies estimate approximately 600–625 genes as significantly associated with CKD onset and progression.^{75,76} The representative signalling network is shown in Figure 2. This summarizes major genetic regulators and interconnected pathways implicated in CKD pathogenesis. While this schematic is not exhaustive, it provides insight into the molecular interplay between inflammation, oxidative stress, fibrotic remodelling, and metabolic dysregulation underlying CKD.

REACTOME PATHWAY

This network diagram illustrates key gene-pathway associations involved in chronic kidney disease (CKD). Although over 600 genes are linked to CKD, the figure highlights representative genes and their roles in critical processes such as podocyte function, RAAS regulation, creatine metabolism, immune response, and cytoskeletal organization. Blue nodes represent genes, while green nodes represent biological functions, with edges indicating gene involvement. The diagram reveals a modular structure,

Table 1: Chromosome specific entries of genes with their total number and association with CKD.⁷⁴

| Chromosome Number | Total Genes on the Chromosomes | Number of Genes Associated with CKD | Names of the Noted CKD Associated Genes |
|-------------------|--------------------------------|-------------------------------------|---|
| 1 | 2,050 | 21 | FAM78B, MYH9, APOL1 |
| 2 | 1,301 | 47 | APOL1 |
| 3 | 1,079 | 18 | FAM78B |
| 4 | 753 | 25 | SHROOM3 |
| 5 | 884 | 19 | COL4A3, COL4A4 |
| 6 | 1,045 | 60 | UMOD, HLA-DRB1 |
| 7 | 992 | 26 | PKD1, PKD2 |
| 8 | 1,021 | 16 | TSC2 |
| 9 | 778 | 8 | FRMD3 |
| 10 | 731 | 14 | COL4A5 |
| 11 | 1,316 | 33 | WT1 |
| 12 | 1,036 | 26 | TTR |
| 13 | 321 | 7 | VHL |
| 14 | 821 | 1 | COL11A1 |
| 15 | 616 | 33 | CASR |
| 16 | 862 | 29 | BBS1 |
| 17 | 1,188 | 24 | UBE2Z |
| 18 | 269 | 13 | SLC12A3 |
| 19 | 1,474 | 9 | SLC7A9 |
| 20 | 543 | 9 | MMP9 |
| 21 | 232 | 2 | HNF1A |
| 22 | 492 | 8 | TSC2 |
| X | 846 | 4 | AVPR2, PAX2 |
| Y | 63 | 52 | 26 genes |

where distinct yet interconnected pathways collectively maintain kidney function. Disruption in these pathways through mutations or gene dysregulation can impair renal processes, leading to CKD. This highlights the multifactorial nature of CKD and the need for a systems-level understanding to uncover therapeutic targets.⁷⁷

HERBAL REMEDIES TARGETING KIDNEY DISEASES AND THEIR PATHWAYS

Herbal therapeutics have shown promising potential in managing various kidney disorders by modulating molecular pathways involved in fibrosis, oxidative stress, inflammation, and electrolyte imbalance. These plant-based interventions act through bioactive compounds such as curcumin, astragalosides, catechins, and salvianolic acids, which influence signalling mechanisms like mTOR, TGF- β , NF- κ B, and

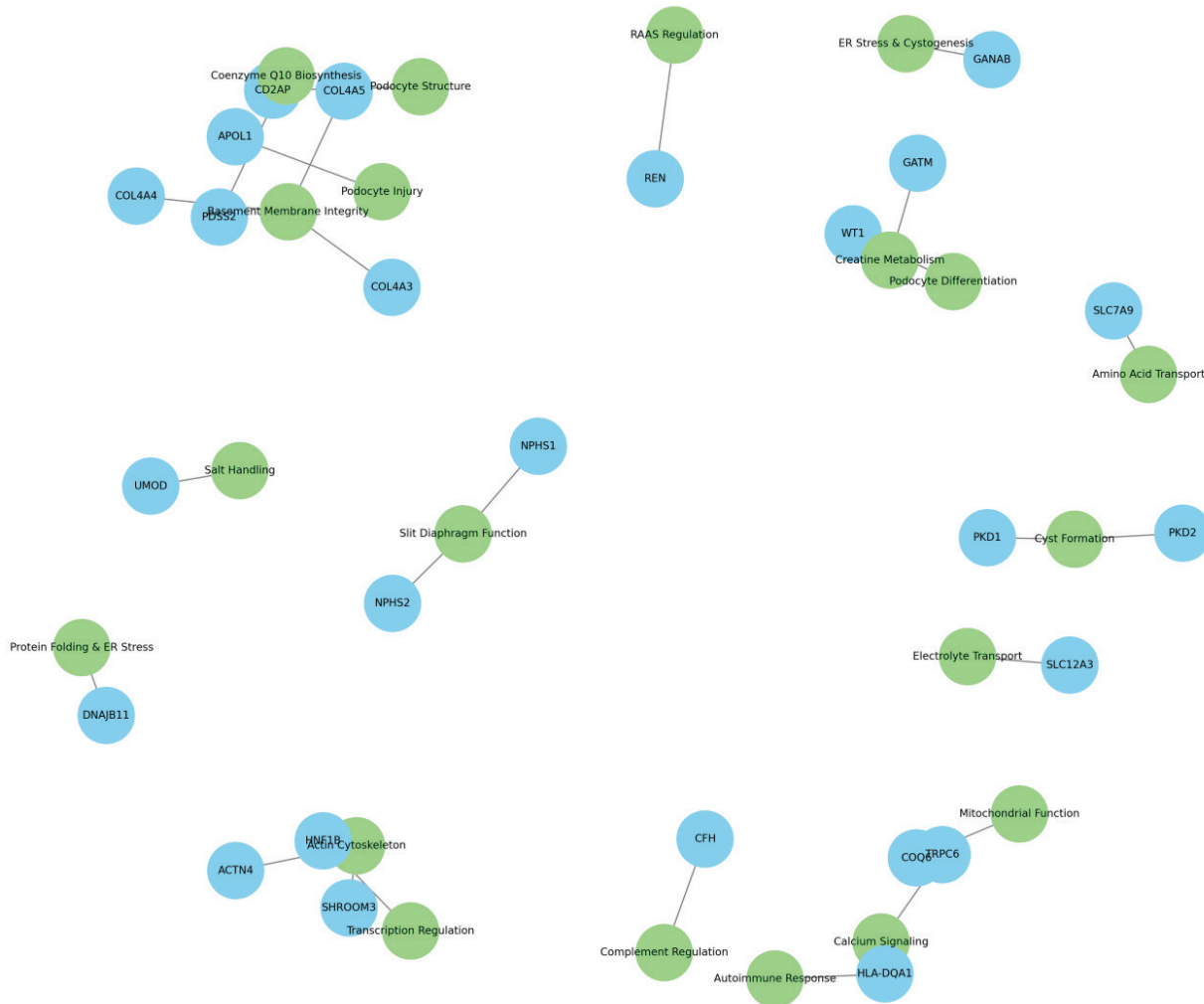


Figure 2: Schematic illustration showing the interplay between key pathways of the genes involved in CKD.

RAAS, thereby slowing disease progression and improving renal function. The following table summarizes kidney conditions, their pathological features, potential herbal remedies, active compounds, and targeted mechanisms.⁷⁸

Herbal Supplements as Sources of Potassium and Phosphorus: Critical Considerations in CKD Management

Potassium and phosphorus play essential roles in cellular function, neuromuscular activity, and energy metabolism; however, their regulation becomes critically important in chronic kidney disease (CKD) management. Impaired renal excretion predisposes CKD patients to hyperkalaemia and hyperphosphatemia, which can cause life-threatening arrhythmias, vascular calcification, bone-mineral disorders, and accelerated cardiovascular risk. Herbal supplements, often perceived as safe, may act as hidden sources of potassium and phosphorus, complicating dietary control strategies that are fundamental in CKD care. Potassium

directly influences RAAS signalling, WNK-SPAK/OSR1 pathways, and genes such as *SLC12A1*, *KCNJ1*, and *WNK1*, which regulate sodium–potassium transport and blood pressure critical targets in CKD. Similarly, phosphorus homeostasis involves the FGF23–Klotho axis, PTH signaling, and Na⁺/Pi cotransporters (*SLC34 family*), linked to genes like *FGF23*, *KL*, and *PTH*, all central to mineral-bone metabolism and cardiovascular protection in CKD patients. Understanding the mineral content of herbal formulations, and their bioactive compounds is essential for safe therapeutic use, preventing electrolyte imbalance while harnessing potential nephroprotective and anti-inflammatory benefits in CKD management.⁹⁹

CKD and Future Prospects

Despite major advancements in understanding CKD pathophysiology, current treatments remain largely supportive, rather than curative. Integrating multi-omics data can deepen insights into monogenic and polygenic

Table 2: Summary of various CKD conditions with their pathological features, herbal interventions, active compounds, and key molecular targets.

| S. No. | CKD Condition | Evidence From Studies | Potential Herbal Drugs | Active Compounds | Targeted Mechanisms |
|--------|---|---|---|---|--|
| 1. | Autosomal Dominant Polycystic Kidney Disease (ADPKD) | Reduces Cyst growth, lowers proliferative indices, induce fibrosis | Curcuma longa, Salvia miltiorrhiza, Resveratrol (Polygonum cuspidatum) Cordyceps sinensis | Curcumin, Salvianolic acids, Resveratrol, Cordycepin | ↓mTOR pathway, ↓TGF-β, anti-inflammatory, antioxidant. ⁷⁹⁻⁸¹ |
| 2. | Autosomal Recessive Polycystic Kidney Disease (ARPKD) | Reduces Hepatorenal cysts, fibrosis, enhances antioxidant activity | Curcuma longa, Astragalus membranaceus, Cordyceps sinensis | Curcumin, Astragaloside IV, Cordycepin | ↓Fibrosis (TGF-β inhibition), antioxidant, mitochondrial support. ^{82,83} |
| 3. | Nephronophthisis (NPHP) | Reduces renal & tubulointerstitial, fibrosis, oxidative stress | Astragalus membranaceus, Curcuma longa, Salvia miltiorrhiza, Rheum officinale | Astragaloside IV, Curcumin, Salvianolic acids, Anthraquinones | ↓TGF-β/Smad signaling, ↓NF-κB, antioxidant. ^{84,85} |
| 4. | Medullary Cystic Kidney Disease (MCKD) | Tubulointerstitial nephritis, hyperuricemia, improved renal function, reduce inflammation | Astragalus membranaceus, Salvia miltiorrhiza, Punica granatum | Astragaloside IV, Salvianolic acids, Polyphenols | ↓Fibrosis, ↓inflammation, improves renal perfusion. ⁸⁶⁻⁸⁸ |
| 5. | Alport Syndrome | Glomerulosclerosis, enhances antioxidant property | Astragalus membranaceus, Curcuma longa, Green Tea, Cordyceps sinensis | Astragaloside IV, Curcumin, Catechins, Cordycepin | ↓Fibrosis, antioxidant, podocyte protection. ^{84,89} |
| 6. | Fabry Disease | reduced oxidative stress markers | Green Tea, Curcuma longa, Salvia miltiorrhiza | Catechins, Curcumin, | Antioxidant, anti-inflammatory. ^{82,90} |
| 7. | Pierson Syndrome | GBM defect, proteinuria | Astragalus membranaceus, Curcuma longa | Astragaloside IV, Curcumin | Podocyte protection, ↓oxidative stress ⁸³ |
| 8. | Congenital Nephrotic Syndrome | attenuated renal injury, Massive proteinuria | Astragalus membranaceus, Curcuma longa | Astragaloside IV, Curcumin | ↓Inflammation, enhances glomerular function. ^{82,87} |
| 9. | Denys-Drash & Frasier Syndromes | Podocyte dysfunction, progressive CKD | Astragalus membranaceus, Salvia miltiorrhiza | Astragaloside IV, Salvianolic acids | Podocyte stabilization, ↓fibrosis. ^{87,91} |
| 10. | Thin Basement Membrane Nephropathy | Mild hematuria | Green Tea, Curcuma longa | Catechins, Curcumin | Antioxidant, GBM protection. ⁹² |
| 11. | Bartter Syndrome | Salt wasting, hypokalemia | Astragalus membranaceus | Astragaloside IV | Electrolyte balance, anti-inflammatory. ⁸⁹ |
| 13. | Cystinuria | ↓ urinary oxalate and uric acid | Phyllanthus niruri, Camellia sinensis | Lignans, Catechins | ↓Stone formation, antioxidant. ⁹³ |
| 14. | Primary Hyperoxaluria | alleviate CaOx crystal deposition in the mouse kidney | Phyllanthus niruri, Curcuma longa | Lignans, Curcumin | ↓Oxalate synthesis, antioxidant. ^{93,94} |
| 17. | Renal Cysts and Diabetes (RCAD) | Cysts | Astragalus membranaceus, Curcuma longa | Astragaloside IV, Curcumin | ↓Fibrosis, antioxidant. ⁹⁵ |
| 20. | Hypertensive Nephropathy | Reduced BP, fibrosis | Curcuma longa, Green Tea | Curcumin, Catechins | ↓BP (RAAS), antioxidant, anti-fibrotic. ^{96,97} |
| 23. | Focal Segmental Glomerulosclerosis (FSGS) | Podocyte loss, sclerosis | Astragalus membranaceus, Salvia miltiorrhiza | Astragaloside IV, Salvianolic acids | Podocyte protection, ↓TGF-β. ^{84,89,98} |
| 24. | Kidney Stone Disease (Nephrolithiasis) | Calcium/uric acid stones | Camellia sinensis, Curcuma longa | Catechins, Curcumin | ↓Crystal aggregation, antioxidant. ⁹⁴ |

Table 3: Herbal supplements serving as sources of potassium and phosphorus, including their common forms, bioactive compounds and consideration for CKD management.^{99,100,101,102,103}

| S. No. | Herb | Common Form | Scientific Name | No. of Bioactive Compounds | P/K Content | CKD Considerations |
|--------|----------------------|-----------------|--------------------------------|----------------------------|----------------|--|
| 1 | Alfalfa | Leaf, Sprout | <i>Medicago sativa</i> | 11 | High K | Risk of hyperkalemia in CKD |
| 2 | Bitter Melon | Fruit, Leaf | <i>Momordica charantia</i> | >60 | Moderate P & K | Hypoglycemia risk |
| 4 | Coriander | Leaf, Seed | <i>Coriandrum sativum</i> | 44–53 | Moderate P & K | Allergy possible |
| 5 | Evening Primrose | Oil, Capsule | <i>Oenothera biennis</i> | 2 | Moderate P | Interaction with anticoagulants |
| 7 | Sugar Kelp | Whole Plant | <i>Saccharina latissima</i> | Several | High K | Avoid in severe CKD |
| 9 | Purslane | Leaf | <i>Portulaca oleracea</i> | 184 | High K | Hyperkalemia risk |
| 10 | Scullcap | Root, Leaf | <i>Scutellaria baicalensis</i> | >300 | Moderate K | Monitor K in CKD |
| 11 | Turmeric | Rhizome, Powder | <i>Curcuma longa</i> | ~235 | Moderate P | Drug interactions |
| 12 | American Ginseng | Root, Extract | <i>Panax quinquefolius</i> | >100 | Moderate P | May alter BP and glucose |
| 13 | Black Mustard | Leaf | <i>Brassica nigra</i> | 26 | Moderate K | Thyroid caution |
| 14 | Chicory | Leaf, Root | <i>Cichorium intybus</i> | 15 | Moderate K | GI discomfort possible |
| 15 | Dandelion | Root, Leaf | <i>Taraxacum officinale</i> | >300 | High K | Hyperkalemia risk |
| 16 | Feverfew | Leaf | <i>Tanacetum parthenium</i> | 39 | Moderate P | Avoid with anticoagulants |
| 17 | Gotu Kola | Leaf | <i>Centella asiatica</i> | Several | Moderate K | Mild diuretic effect; monitor K |
| 18 | Kudzu | Root, Shoot | <i>Pueraria tuberosa</i> | 23 | Moderate K | Phytoestrogen activity caution |
| 19 | Noni | Fruit, Leaf | <i>Morinda citrifolia</i> | 200 | Moderate K | Limited CKD safety evidence |
| 21 | Shepherd's Purse | Whole Plant | <i>Capsella bursa-pastoris</i> | 50 | Moderate P & K | May lower BP; monitor electrolyte balance |
| 22 | Water Lotus | Whole Plant | <i>Nymphaea lotus</i> | 50 | Moderate P | Limited data; avoid overdose |
| 23 | Bai Zhi | Root | <i>Angelica dahurica</i> | >300 | Moderate K | Possible herbal-drug interactions |
| 26 | Dulse | Seaweed | <i>Palmaria palmata</i> | 50 | High K | Avoid in advanced CKD |
| 27 | Garlic | Leaf, Bulb | <i>Allium sativum</i> | 50 | Moderate P | Anticoagulant effect; BP lowering |
| 28 | Japanese Honeysuckle | Flower | <i>Lonicera japonica</i> | 140 | Moderate K | Limited CKD data |
| 29 | Lemongrass | Leaf | <i>Cymbopogon citratus</i> | 70 | Moderate K | Monitor K in CKD |
| 30 | Papaya | Leaf, Fruit | <i>Carica papaya</i> | 40 | Moderate K | May affect glucose regulation |
| 32 | Stinging Nettle | Leaf | <i>Urtica dioica</i> | 40 | Moderate P & K | Diuretic effect; monitor electrolytes |
| 33 | Borage | Leaf, Capsule | <i>Borago officinalis</i> | 20–25 | Moderate P | Possible anticoagulant interactions |
| 35 | Flaxseed | Seed, Oil | <i>Linum usitatissimum</i> | 40+ | Moderate P | High fiber; digestive discomfort possible |
| 38 | Milk Thistle | Seed, Extract | <i>Silybum marianum</i> | ~200 | Moderate P | Nephroprotective; drug interactions possible |
| 39 | Onion | Leaf | <i>Allium cepa</i> | 40+ | Moderate P | Anticoagulant interaction; allergy risk |
| 42 | Sunflower | Seed, Oil | <i>Helianthus annuus</i> | ~30 | Moderate P | High calories; avoid in obese CKD |
| 43 | Yellow Dock | Root, Capsule | <i>Rumex crispus</i> | 20+ | Moderate P | GI irritation possible; avoid excess intake |

forms, while precision medicine, CRISPR gene editing, and AI-driven approaches hold promise for targeted interventions and early risk prediction. In parallel, the exploration of herbal therapeutics offers a promising alternative to conventional treatment regimens. However, meticulous translational studies are needed to validate the molecular mechanisms, pharmacokinetics, and safety profiles of plant-derived compounds, as well as to standardize their formulations.

CONCLUSION

In the search for reliable and safe herbal formulations to manage CKD, we explored key genes involved in renal function, their chromosomal localization, and their regulatory roles. Under normal physiological conditions, these genes contribute to maintaining kidney homeostasis. However, in pathological states, their dysregulation leads to cellular dysfunction and progression of CKD. While several genetic and environmental contributors have been identified, many remain undiscovered. This review highlights existing and investigational drug molecules that have shown therapeutic potential in CKD, with some already validated through multiple clinical trials. Herbal formulations, due to their multifaceted bioactivity and historical usage, offer a promising complementary approach to current treatments. If properly characterized and standardized, these formulations may progress toward regulatory approval and clinical use. However, the safety profile of herbal medicines must be carefully assessed. Herbal therapies hold great promise, rigorous identification, validation, and standardization are essential. Alternatively, these formulations could be developed as dietary supplements to support renal health. We hope this report will encourage further research and evoke clinical interest in evidence-based herbal therapeutics for CKD, promoting safer, accessible, and culturally relevant treatment strategies.

ACKNOWLEDGMENT

Authors are grateful to Professor Abbas Ali Mahdi, VC, Era University for his excellent administrative support and continued academic indulgence.

REFERENCES

- Bikbov B, Purcell CA, Levey AS, Smith M, Abdoli A, Abebe M, Adebayo OM, Afarideh M, Agarwal SK, Agudelo-Botero M, Ahmadian E. Global, regional, and national burden of chronic kidney disease, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. *The Lancet*. 2020 Feb 29;395(10225):709–33.
- Luyckx VA, Tonelli M, Stanifer JW. The global burden of kidney disease and the sustainable development goals. *Bulletin of the World Health Organization*. 2018 Apr 20;96(6):414.
- Shabaka A, Cases-Corona C, Fernandez-Juarez G. Therapeutic insights in chronic kidney disease progression. *Frontiers in medicine*. 2021 Feb 23;8:645187.
- Sharma K, et al. Novel signaling pathways in CKD: oxidative stress and beyond. *Kidney Int*. 2021;99(3):593–607.
- Pattaro C, et al. Genetic associations at 53 loci highlight cell types and biological pathways relevant for kidney function. *Nat Commun*. 2016;7:10023.
- Wuttke M, Köttgen A. Genetic studies of kidney disease: a new era. *Nat Rev Nephrol*. 2016;12(10):610–625.
- Köttgen A, et al. Uromodulin and kidney function: insights from genetic and proteomic studies. *Nat Rev Nephrol*. 2020;16(10):567–581.
- Freedman BI, et al. APOL1 gene variants in CKD and ESRD. *Nat Rev Nephrol*. 2018;14(10):614–626.
- Park S, et al. RAAS gene polymorphisms and kidney disease risk: meta-analysis. *PLoS One*. 2021;16(7):e0254318.
- Wing MR, et al. Epigenetics in CKD progression. *Nat Rev Nephrol*. 2022;18(1):43–58.
- Heerspink HJL, et al. SGLT2 inhibitors in kidney disease. *Lancet Diabetes Endocrinol*. 2020;8(7):605–615.
- Bakris GL, et al. Adverse events of RAAS blockers in CKD. *Kidney Int Suppl*. 2021;11(1):25–33.
- Wang Y, et al. Traditional herbal medicine for CKD. *Front Pharmacol*. 2022;13:815888.
- Aggarwal BB, Harikumar KB. Potential therapeutic effects of curcumin, the anti-inflammatory agent, against CKD. *Br J Pharmacol*. 2009;165(5):1361–1376.
- Zhang Y, et al. Astragalus membranaceus and kidney protection: molecular mechanisms. *Am J Chin Med*. 2020;48(7):1599–1617.
- Kong WJ, et al. Berberine: AMPK activation and kidney health. *Metabolism*. 2019;102:153983.
- Posadzki P, et al. Herb-drug interactions in CKD: clinical implications. *Br J Clin Pharmacol*. 2018;84(4):679–690.
- Torres VE, Harris PC, Pirson Y. Autosomal dominant polycystic kidney disease. *Lancet*. 2007;369(9569):1287–1301.
- Vasileva VY, Sultanova RF, Sudarikova AV, Ilatovskaya DV. Insights Into the Molecular Mechanisms of Polycystic Kidney Diseases. *Front Physiol*. 2021 Sep 8;12:693130. doi: 10.3389/fphys.2021.693130. PMID: 34566674; PMCID: PMC8456103.
- Wilson PD. Polycystic kidney disease. *N Engl J Med*. 2004;350:151–164.
- Calvet JP. The role of calcium and cyclic AMP in PKD. *Exon Publications*. 2015 Nov 18:169–96.
- Shillingford JM, et al. The mTOR pathway is regulated by polycystin-1, and its inhibition reverses renal cystogenesis in polycystic kidney disease. *PNAS*. 2006;103(14):5466–5471.
- Bergmann C, Guay-Woodford LM, Harris PC, Horie S, Peters DJM, Torres VE. Polycystic kidney disease. *Nat Rev Dis Primers*. 2018 Dec 6;4(1):50. doi: 10.1038/s41572-018-0047-y. PMID: 30523303; PMCID: PMC6592047.
- Ward CJ, Hogan MC, Rossetti S, Walker D, Sneddon T, Wang X, Kubly V, Cunningham JM, Bacallao R, Ishibashi M, Milliner DS. The gene mutated in autosomal recessive polycystic kidney disease encodes a large, receptor-like protein. *Nature genetics*. 2002 Mar;30(3):259–69.
- Hildebrandt F, Benzing T, Katsanis N. Ciliopathies. *New England Journal of Medicine*. 2011 Apr 21;364(16):1533–43.

26. Zhang MZ, Mai W, Li C, Cho SY, Hao C, Moeckel G, Zhao R, Kim I, Wang J, Xiong H, Wang H. PKHD1 protein encoded by the gene for autosomal recessive polycystic kidney disease associates with basal bodies and primary cilia in renal epithelial cells. *Proceedings of the National Academy of Sciences*. 2004 Feb 24;101(8):2311-6.
27. Dalagiorou G, Basdra EK, Papavassiliou AG. Polycystin-1: function as a mechanosensor. *The international journal of biochemistry & cell biology*. 2010 Oct 1;42(10):1610-3.
28. Rossetti S, Consugar MB, Chapman AB, Torres VE, Guay-Woodford LM, Grantham JJ, Bennett WM, Meyers CM, Walker DL, Bae K, Zhang QJ. Comprehensive molecular diagnostics in autosomal dominant polycystic kidney disease. *Journal of the American Society of Nephrology*. 2007 Jul 1;18(7):2143-60.
29. Wolf MT, Bonsib SM, Larsen CP, Hildebrandt F. Nephronophthisis: a pathological and genetic perspective. *Pediatric Nephrology*. 2024 Jul;39(7):1977-2000.
30. Fliegau M, Horvath J, von Schnakenburg C, Olbrich H, Müller D, Thumfart J, Schermer B, Pazour GJ, Neumann HP, Zentgraf H, Benzing T, Omran H. Nephrocystin specifically localizes to the transition zone of renal and respiratory cilia and photoreceptor connecting cilia. *J Am Soc Nephrol*. 2006 Sep;17(9):2424-33. doi: 10.1681/ASN.2005121351. Epub 2006 Aug 2. PMID: 16885411.
31. Hynes AM, Giles RH, Srivastava S, Eley L, Whitehead J, Danilenko M, Raman S, Slaats GG, Colville JG, Ajzenberg H, Kroes HY, Thelwall PE, Simmons NL, Miles CG, Sayer JA. Murine Joubert syndrome reveals Hedgehog signaling defects as a potential therapeutic target for nephronophthisis. *Proc Natl Acad Sci U S A*. 2014 Jul 8;111(27):9893-8. doi: 10.1073/pnas.1322373111. Epub 2014 Jun 19. PMID: 24946806; PMCID: PMC4103340.
32. Bleyer AJ, Kmoch S. Autosomal dominant tubulointerstitial kidney disease: of names and genes. *Kidney Int*. 2014 Sep;86(3):459-61. doi: 10.1038/ki.2014.125. PMID: 25168494.
33. Pottel H, Delanaye P, Schaeffner E, Dubourg L, Eriksen BO, Melsom T, Lamb EJ, Rule AD, Turner ST, Glassock RJ, De Souza V. Estimating glomerular filtration rate for the full age spectrum from serum creatinine and cystatin C. *Nephrology Dialysis Transplantation*. 2017 Mar 1;32(3):497-507.
34. Thorner PS. Alport syndrome and thin basement membrane nephropathy. *Nephron Clinical practice*. 2007 Jun 6;106(2):c82-8.
35. Kashtan, C. E. (2021). Alport Syndrome: Facts and Opinions. *F1000Research*, 10, 28.
36. Zhu H, Liao J, Zhou X, Hong X, Song D, Hou FF, Liu Y, Fu H. Tenascin-C promotes acute kidney injury to chronic kidney disease progression by impairing tubular integrity via $\alpha v \beta 6$ integrin signaling. *Kidney international*. 2020 May 1;97(5):1017-31.
37. Desnick, R. J., Ioannou, Y. A., & Eng, C. M. (2001). α -Galactosidase A deficiency: Fabry disease. In *The Metabolic and Molecular Bases of Inherited Disease*.
38. Aerts JM, Groener JE, Kuiper S, Donker-Koopman WE, Strijland A, Ottenhoff R, van Roomen C, Mirzaian M, Wijburg FA, Linthorst GE, Vedder AC. Elevated globotriaosylsphingosine is a hallmark of Fabry disease. *Proceedings of the National Academy of Sciences*. 2008 Feb 26;105(8):2812-7.
39. Brady RO, Schiffmann R. Clinical features of and recent advances in therapy for Fabry disease. *Jama*. 2000 Dec 6;284(21):2771-5.
40. Beaufils C, Farlay D, Machuca-Gayet I, Fassier A, Zenker M, Freychet C, Bonnelye E, Bertholet-Thomas A, Ranchin B, Bacchetta J. Skeletal impairment in Pierson syndrome: Is there a role for laminin $\beta 2$ in bone physiology?. *Bone*. 2018 Jan 1;106:187-93.
41. Lipska-Ziętkiewicz BS, Ozaltin F, Hölttä T, Bockenhauer D, Bérody S, Levchenko E, Vivarelli M, Webb H, Haffner D, Schaefer F, Boyer O. Genetic aspects of congenital nephrotic syndrome: a consensus statement from the ERKNet-ESPN inherited glomerulopathy working group. *European Journal of Human Genetics*. 2020 Oct;28(10):1368-78.
42. Jalanko, H. (2009). Congenital nephrotic syndrome. *Pediatric Nephrology*, 24(11), 2121-2128.
43. Barbaux S, Niaudet P, Gubler MC, Grünfeld JP, Jaubert F, Kuttent F, Fékété CN, Souleyreau-Therville N, Thibaud E, Fellous M, McElreavey K. Donor splice-site mutations in WT1 are responsible for Frasier syndrome. *Nature genetics*. 1997 Dec 1;17(4):467-70.
44. Yaqing CA, Baocheng GU, Min NI. Genotype-Phenotype Correlation Analysis of WT1 Gene Variants in Denys-Drash Syndrome and Frasier Syndrome. *Journal of Rare Diseases*. 2024 Jan 30;3(1):63-76.
45. Kashtan, C. E. (2021). Alport Syndrome and Thin Basement Membrane Nephropathy: Diseases of the Glomerular Basement Membrane. *Kidney International*, 100(4), 867-878.
46. Wang YY, Rana K, Tonna S, Lin T, Sin L, Savige J. COL4A3 mutations and their clinical consequences in thin basement membrane nephropathy (TBMN). *Kidney international*. 2004 Mar 1;65(3):786-90.
47. Weber S, Strasser K, Rath S, Kittke A, Beicht S, Alberer M, Lange-Sperandio B, Hoyer PF, Benz MR, Ponsel S, Weber LT. Identification of 47 novel mutations in patients with Alport syndrome and thin basement membrane nephropathy. *Pediatric Nephrology*. 2016 Jun;31(6):941-55.
48. Seyberth, H. W., & Schlingmann, K. P. (2011). Bartter- and Gitelman-like syndromes: salt-losing tubulopathies with loop or DCT defects. *Pediatric Nephrology*, 26(10), 1789-1802.
49. Takemori S, Tanigaki S, Nozu K, Yoshihashi H, Uchiumi Y, Sakaguchi K, Tsushima K, Kitamura A, Kobayashi C, Matsuhima M, Tajima A. Prenatal diagnosis of MAGED2 gene mutation causing transient antenatal Bartter syndrome. *European Journal of Medical Genetics*. 2021 Oct 1;64(10):104308.
50. Palazzo V, Raglianti V, Landini S, Cirillo L, Errichiello C, Buti E, Artuso R, Tiberi L, Vergani D, Dirupo E, Romagnani P. Clinical and genetic characterization of patients with bartter and gitelman syndrome. *International Journal of Molecular Sciences*. 2022 May 18;23(10):5641.
51. Simon, D. B., et al. (1996). Gitelman's variant of Bartter's syndrome, inherited hypokalaemic alkalosis, is caused by mutations in the thiazide-sensitive Na-Cl cotransporter. *Nature Genetics*, 12, 24-30.
52. Viering D, Schlingmann KP, Hureauux M, Nijenhuis T, Mallett A, Chan MMY, van Beek A, van Eerde AM, Coulibaly JM, Vallet M, Decramer S, Pelletier S, Klaus G, Kömhoff M, Beetz R, Patel C, Shenoy M, Steenbergen EJ, Anderson G, Bongers EMHF, Bergmann C, Panneman D, Rodenburg RJ, Kleta R, Houillier P, Konrad M, Vargas-Poussou R, Knoers NVAM, Bockenhauer D, de Baaij JHF; Genomics England Research Consortium. Gitelman-Like Syndrome Caused by Pathogenic Variants in mtDNA. *J Am Soc Nephrol*. 2022

- Feb;33(2):305-325. doi: 10.1681/ASN.2021050596. Epub 2021 Oct 4. PMID: 34607911; PMCID: PMC8819995.
53. Eggermann, T., Venghaus, A., Zerres, K. (2012). Cystinuria: an inborn cause of urolithiasis. *Orphanet Journal of Rare Diseases*, 7, 19.
 54. Calonge, M. J., Gasparini, P., Chillarón, J., et al. (1994). Cystinuria caused by mutations in rBAT, a gene involved in the transport of cystine. *Nature Genetics*, 6, 420–425.
 55. Font-Llitjós, M., Jiménez-Vidal, M., Bisceglia, L., et al. (2005). New insights into cystinuria: 40 new mutations, genotype-phenotype correlation, and digenic inheritance causing cystinuria. *Human Mutation*, 26(5), 415–421.
 56. Tanase DM, Gosav EM, Anton MI, Floria M, Seritean Isac PN, Hurjui LL, Tarniceriu CC, Costea CF, Ciocoiu M, Rezus C. Oxidative stress and NRF2/KEAP1/ARE pathway in diabetic kidney disease (DKD): new perspectives. *Biomolecules*. 2022 Sep 2;12(9):1227.
 57. Köttgen, A., et al. (2009). Multiple loci associated with indices of renal function and chronic kidney disease. *Nat Genet*, 41(6), 712–717.
 58. Khan Z, Pandey M. Role of kidney biomarkers of chronic kidney disease: An update. *Saudi journal of biological sciences*. 2014 Sep 1;21(4):294-9.
 59. Kato, M., & Natarajan, R. (2019). Epigenetics and epigenomics in diabetic kidney disease and metabolic memory. *Nature Reviews Nephrology*, 15(6), 327–345.
 60. Pagtalunan, M. E., et al. (1997). Structural-functional relationships in diabetic nephropathy: role of the glomerular basement membrane. *Journal of Clinical Investigation*, 99(10), 2470–2478.
 61. Geng XD, Wang WW, Feng Z, Liu R, Cheng XL, Shen WJ, Dong ZY, Cai GY, Chen XM, Hong Q, Wu D. Identification of key genes and pathways in diabetic nephropathy
 62. Peng Z, Xu Q, Hu W, Cheng Y. Review on molecular mechanism of hypertensive nephropathy. *Current Pharmaceutical Design*. 2023 Sep 1;29(32):2568-78.
 63. Liu T, Liu M, Shang P, Jin X, Liu W, Zhang Y, Li X, Ding Y, Li Y, Wen A. Investigation into the underlying molecular mechanisms of hypertensive nephrosclerosis using bioinformatics analyses. *Mol Med Rep*. 2018 Mar;17(3):4440-4448. doi: 10.3892/mmr.2018.8405. Epub 2018 Jan 9. PMID: 29328390; PMCID: PMC5802219.
 64. Zubirán NS. Urinary Serpin-A3 is an early predictor of clinical response to therapy in patients with 1 proliferative Lupus Nephritis 2.
 65. Tsokos GC. Autoimmunity and organ damage in systemic lupus erythematosus. *Nat Immunol*. 2020;21(6):605-614.
 66. Cheung CK, Alexander S, Reich HN, Selvaskandan H, Zhang H, Barratt J. The pathogenesis of IgA nephropathy and implications for treatment. *Nature Reviews Nephrology*. 2025 Jan;21(1):9-23.
 67. Khan SR, Pearle MS, Robertson WG, et al. Kidney stones. *Nat Rev Dis Primers*. 2021;7(1):41.
 68. Howles SA, Wiberg A, Goldsworthy M, et al. Genetic determinants of kidney stone disease. *Nat Commun*. 2022;13:1953.
 69. Singh, P., Harris, P.C., Sas, D.J. et al. The genetics of kidney stone disease and nephrocalcinosis. *Nat Rev Nephrol* 18, 224–240 (2022). <https://doi.org/10.1038/s41581-021-00513-4>
 70. Angulo JC, Manini C, López JI, Pueyo A, Colás B, Ropero S. The role of epigenetics in the progression of clear cell renal cell carcinoma and the basis for future epigenetic treatments. *Cancers*. 2021 Apr 25;13(9):2071.
 71. Hsieh, J. J., Purdue, M. P., Signoretti, S., et al. (2017). Renal cell carcinoma. *Nature Reviews Disease Primers*, 3, 17009.
 72. Turajlic, S., Xu, H., Litchfield, K., et al. (2018). Tracking cancer evolution reveals constrained routes to metastases: TRACERx Renal. *Cell*, 173(3), 581–594.e12.
 73. Motzer, R. J., Jonasch, E., Agarwal, N., et al. (2021). Kidney cancer, version 3.2022, NCCN Clinical Practice Guidelines in Oncology. *Journal of the National Comprehensive Cancer Network*, 19(1), 19–37.
 74. OMIM database: Online Mendelian Inheritance in Man (OMIM). <https://www.omim.org>.
 75. Wuttke, M., Li, Y., Li, M., et al. (2019). A catalog of genetic loci associated with kidney function from analyses of a million individuals. *Nature Genetics*, 51(6), 957–972.
 76. Stanzick, K. J., Li, Y., Schlosser, P., et al. (2021). Discovery and prioritization of variants and genes for kidney function in >1.2 million individuals. *Nature Communications*, 12(1), 4350.
 77. Kegg Network Database. <https://www.genome.jp/kegg/network.html>. Accessed on 04, Aug, 2025.
 78. Borkar P, Yadav V, Tiwari RR, Samarth RM. A systematic review of potential candidates of herbal medicine in treatment of chronic kidney disease. *Phytomedicine Plus*. 2022 Nov 1;2(4):100361.
 79. Li Y, Gao J, Yang X, Li T, Yang B, Aili A. Combination of curcumin and ginkgolide B inhibits cystogenesis by regulating multiple signaling pathways. *Mol Med Rep*. 2021 Mar;23(3):195. doi: 10.3892/mmr.2021.11834. Epub 2021 Jan 26. PMID: 33495815; PMCID: PMC7821343.
 80. Gao J, Zhou H, Lei T, Zhou L, Li W, Li X, Yang B. Curcumin inhibits renal cyst formation and enlargement in vitro by regulating intracellular signaling pathways. *European journal of pharmacology*. 2011 Mar 1;654(1):92-9.
 81. Wu M, Gu J, Mei S, Xu D, Jing Y, Yao Q, Chen M, Yang M, Chen S, Yang B, Qi N, Hu H, Wüthrich RP, Mei C. Resveratrol delays polycystic kidney disease progression through attenuation of nuclear factor κB-induced inflammation. *Nephrol Dial Transplant*. 2016 Nov;31(11):1826-1834. doi: 10.1093/ndt/gfw058. Epub 2016 Apr 19. PMID: 27190325.
 82. Gaedeke J, Noble NA, Border WA. Curcumin blocks multiple sites of the TGF-beta signaling cascade in renal cells. *Kidney Int*. 2004 Jul;66(1):112-20. doi: 10.1111/j.1523-1755.2004.00713.x. PMID: 15200418.
 83. Xu W, Shao X, Tian L, Gu L, Zhang M, Wang Q, Wu B, Wang L, Yao J, Xu X, Mou S, Ni Z. Astragaloside IV ameliorates renal fibrosis via the inhibition of mitogen-activated protein kinases and antiapoptosis in vivo and in vitro. *J Pharmacol Exp Ther*. 2014 Sep;350(3):552-62. doi: 10.1124/jpet.114.214205. Epub 2014 Jun 20. PMID: 24951279.
 84. Wang L, Chi YF, Yuan ZT, Zhou WC, Yin PH, Zhang XM, Peng W, Cai H. Astragaloside IV inhibits renal tubulointerstitial fibrosis by blocking TGF-β/Smad signaling pathway in vivo and in vitro. *Exp Biol Med (Maywood)*. 2014 Oct;239(10):1310-24. doi: 10.1177/1535370214532597. Epub 2014 May 30. PMID: 24879422.
 85. Lin P, Qiu F, Wu M, Xu L, Huang D, Wang C, Yang X, Ye C. Salvianolic acid B attenuates tubulointerstitial fibrosis by inhibiting EZH2 to regulate the PTEN/Akt pathway. *Pharm Biol*. 2023 Dec;61(1):23-29. doi: 10.1080/13880209.2022.2148169. PMID: 36524761; PMCID: PMC9762854.

86. Murata I, Abe Y, Yaginuma Y, Yodo K, Kamakari Y, Miyazaki Y, Baba D, Shinoda Y, Iwasaki T, Takahashi K, Kobayashi J, Inoue Y, Kanamoto I. Astragaloside-IV prevents acute kidney injury and inflammation by normalizing muscular mitochondrial function associated with a nitric oxide protective mechanism in crush syndrome rats. *Ann Intensive Care*. 2017 Sep 4;7(1):90. doi: 10.1186/s13613-017-0313-2. PMID: 28871521; PMCID: PMC5583140.
87. Zhang D, Li Z, Gao Y, Sun H. Astragaloside IV improves renal function and alleviates renal damage and inflammation in rats with chronic glomerulonephritis. *Turk J Biol*. 2022 Dec 9;47(1):61-73. doi: 10.55730/1300-0152.2641. PMID: 37529109; PMCID: PMC10387845.
88. Radajewska A, Szyller J, Niewiadomska J, Noszczyk-Nowak A, Bil-Lula I. Punica granatum L. polyphenolic extract as an antioxidant to prevent kidney injury in metabolic syndrome rats. *Oxidative Medicine and Cellular Longevity*. 2023;2023(1):6144967.
89. Hu Z, Zhou Y, Gao C, Liu J, Pan C, Guo J. Astragaloside IV attenuates podocyte apoptosis via regulating TXNIP/NLRP3/GSDMD signaling pathway in diabetic nephropathy. *Diabetology & Metabolic Syndrome*. 2024 Dec 18;16(1):296.
90. Bertoldi G, Carraro G, Ravarotto V, Di Vico V, Baldini Anastasio P, Vitturi N, Francini F, Stefanelli LF, Calò LA. The Effect of Green Tea as an Adjuvant to Enzyme Replacement Therapy on Oxidative Stress in Fabry Disease: A Pilot Study. *Front Nutr*. 2022 Jul 8;9:924710. doi: 10.3389/fnut.2022.924710. PMID: 35873439; PMCID: PMC9304972.
91. He Y, Lu R, Wu J, Pang Y, Li J, Chen J, Liu B, Zhou Y, Zhou J. Salvianolic acid B attenuates epithelial-mesenchymal transition in renal fibrosis rats through activating Sirt1-mediated autophagy. *Biomedicine & Pharmacotherapy*. 2020 Aug 1;128:110241.
92. Barocio-Pantoja M, Quezada-Fernández P, Cardona-Müller D, Jiménez-Cázar MB, Larios-Cárdenas M, González-Radillo OI, García-Sánchez A, Carmona-Huerta J, Chávez-Guzmán AN, Díaz-Preciado PA, Balleza-Alejandri R, Pascoe-González S, Grover-Páez F. Green Tea Extract Increases Soluble RAGE and Improves Renal Function in Patients with Diabetic Nephropathy. *J Med Food*. 2021 Dec;24(12):1264-1270. doi: 10.1089/jmf.2020.0212. Epub 2021 Nov 17. PMID: 34788550.
93. Pucci ND, Marchini GS, Mazzucchi E, Reis ST, Srougi M, Evazian D, Nahas WC. Effect of phyllanthus niruri on metabolic parameters of patients with kidney stone: a perspective for disease prevention. *Int Braz J Urol*. 2018 Jul-Aug;44(4):758-764. doi: 10.1590/S1677-5538.IBJU.2017.0521. PMID: 29617079; PMCID: PMC6092661.
94. Li Y, Zhang J, Liu H, Yuan J, Yin Y, Wang T, Cheng B, Sun S, Guo Z. Curcumin ameliorates glyoxylate-induced calcium oxalate deposition and renal injuries in mice. *Phytomedicine*. 2019 Aug 1;61:152861.
95. Sun H, Wang W, Han P, Shao M, Song G, Du H, Yi T, Li S. Astragaloside IV ameliorates renal injury in db/db mice. *Scientific reports*. 2016 Sep 2;6(1):32545.
96. Li C, Chen X, Yao J, Zha W, Li M, Shen J, Jiang H, Tian P. Curcumin modulated gut microbiota and alleviated renal fibrosis in 5/6 nephrectomy-induced chronic kidney disease rats. *PloS one*. 2025 Jan 9;20(1):e0314029.
97. Kanlaya R, Thongboonkerd V. Protective Effects of Epigallocatechin-3-Gallate from Green Tea in Various Kidney Diseases. *Adv Nutr*. 2019 Jan 1;10(1):112-121. doi: 10.1093/advances/nmy077. PMID: 30615092; PMCID: PMC6370267.
98. Fan, HY., Yang, MY., Qi, D. *et al.* Salvianolic acid A as a multifunctional agent ameliorates doxorubicin-induced nephropathy in rats. *Sci Rep* 5, 12273 (2015). <https://doi.org/10.1038/srep12273>
99. National Kidney Foundation. Herbal supplements and kidney disease [Internet]. 2024 [cited 2025 Jul 25]. Available from: <https://www.kidney.org/kidney-topics/herbal-supplements-and-kidney-disease>
100. American Association of Kidney Patients (AAKP). High potassium in chronic kidney disease: causes, consequences and corrections [Internet]. 2024 [cited 2025 Jul 25]. Available from: <https://aakp.org/high-potassium-in-chronic-kidney-disease-causes-consequences-and-corrections>
101. Kalantar-Zadeh K, Fouque D. Nutritional management of chronic kidney disease. *N Engl J Med*. 2021;384:1769–1779. doi:10.1056/NEJMra2030037
102. Chan L, et al. Potassium intake and chronic kidney disease: a review of evidence and guidelines. *Adv Chronic Kidney Dis*. 2023;30(4):1-8. doi:10.1053/j.ackd.2023.04.007
103. Moe SM, Zidehsarai MP, et al. Vegetarianism, herbal supplements, and kidney disease: phosphorus and potassium considerations. *Clin J Am Soc Nephrol*. 2011;6(2):353–359. doi:10.2215/CJN.05040610